

CHIRONOMIDAE IN FRESHWATER HABITATS IN TENERIFE, CANARY ISLANDS

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INTRODUCTION

Some groups of the freshwater fauna of the Canary Islands have been well studied. These include beetles, blackflies, mosquitoes, odonates and water bugs (see Malmqvist *et al.*, 1993 for a review of studies). The Chironomidae of the Canary Islands in contrast have been the subject of only occasional investigations (Becker, 1908; Santos Abreu, 1918; Storå, 1936). Subsequent collections of adult material have been made (Armitage, 1987; Armitage and Tuiskunen, 1988) and Cranston and Armitage (1988) redescribed material from the earlier collections of Becker and Santos Abreu. More recently 7 permanent freshwater streams on the island of Tenerife have been examined by Malmqvist *et al.* (1993). Chironomid data are presented in that paper and this present study analyses those records together with new information from a number of additional sites, based largely on larval material.

STUDY AREA AND METHODS

Tenerife is the largest (2057km²) and highest (3718m) of the Canary Islands and is situated at latitude 28°15'N, 300km off the coast of Morocco in the trade wind belt. The island has a volcanic origin and has never been connected to the African mainland (Schmincke, 1976). This isolation together with the arid nature of the southern part of the island which lies within a rain shadow result in a paucity of freshwater fauna and natural freshwater habitats. This situation is exacerbated by the continuing pressure on existing water sources.

The habitats sampled included stream riffles (7 localities on 'permanent' streams), stream pools (11 localities), streams general, that is not specifically pool or riffle samples (17 localities), residual pools in dried-up stream

beds (12 localities), 4 ponds, 4 reservoirs, 14 artificial pipes, channels ('canals') and 15 seepages/trickles (wet walls). The location of the sites is indicated in Fig. 1 and the altitudinal distribution is illustrated in Fig. 2. Some localities were sampled on more than one occasion in 'spring' and 'autumn/winter' in 1991 and 1993 and details are presented in the appendix. Details on sampling methodology and information on the physical and chemical features of the main stream sites are presented in Malmqvist *et al.* (1993).

The majority of material was larval hence most records are at genus level but the inclusion of some pupae and adults facilitated identification to species in some cases. Light traps located by three streams also provided some adult material. Semi-quantitative data on spatial and temporal changes in the chironomid communities of the 7 'permanent' streams are presented in

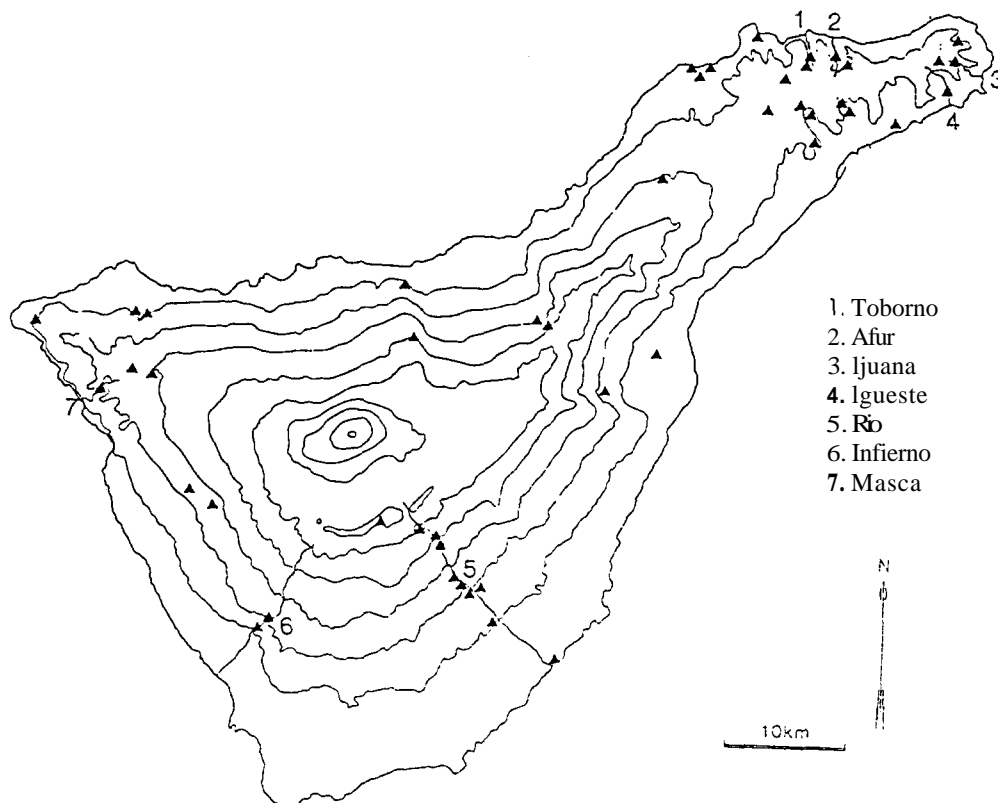


Figure 1. Tenerife; location of sampling locations in relation to the 7 main stream sites.

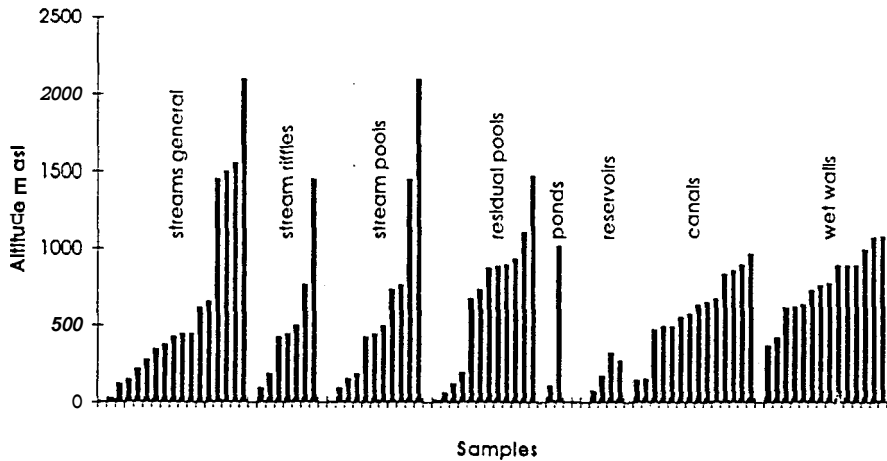


Figure 2. The altitudinal distribution of sample sites per habitat type.

Malmqvist *et al.* (1993) but in this present work most samples were not quantitative and to ensure compatibility all are treated qualitatively.

The data from the collections were analysed to examine the relationship between the chironomid community as a whole to the particular habitat types using ordination techniques. In addition the richness of habitat types in relation to sample effort is examined briefly.

RESULTS

The distribution of taxa in the habitat types is presented in Table 1. Forty-three separate taxa were recorded but the list includes some taxa groups and larvae unidentifiable to species. Of the species, most are cosmopolitan or European in distribution and the African element is weak.

The most commonly occurring taxa in stream pools were *Ablabesmyia longistyla* Fittkau, *Zarelimyia nubila* (Meigen) and *Chironomus* spp.. In riffles *Rheotanytarsus* spp. and *Thienemaniella clavicornis* (Kieffer) were the common forms and in residual pools, ponds and reservoirs *Chironomus* spp. dominated the records. In 'canals' *Paratrichocladius* spp. (probably *rufiventris* (Meigen)) with *Rheotanytarsus* spp. were the most commonly occurring taxa. Chironomidae were seldom

found in 'wet wall' samples and here the most common taxa were *Rheocricotopus* sp, *Paratrichocladius* sp. and *O. (Eudactylocladius)* sp.

The association of assemblages of chironomid taxa with habitat type was tested using Detrended Correspondence Analysis [DCA] (DECORANA; Hill, 1979) in the program CANOCO (ter Braak, 1988). The data matrix consisted of a reduced list of 411 taxa (see Fig. 3) at 118 sites. Ordination resulting from DCA, groups samples on the basis of their faunal composition and the relationship between axis scores and environmental variables allows the detection of general trends governing the distribution of samples. A preliminary run showed that the taxon *Halocladus*, which occurred at only one site not associated with any other species, was exerting a distorting effect on the ordination and it was omitted from the list of taxa. Fig. 3a presents the results of the site ordination. Clearly there is much variation and considerable overlap between habitat types. Nevertheless, lentic and lotic components are recognizable. The ordination of taxa is shown in Fig. 3b. and reflects the same lentic-lotic trend from right to left along axis 1, but no well-defined assemblages of taxa could be detected in response to underlying environmental gradients, a feature which may be linked to the opportunistic response of chironomid species to habitat availability (Rossaro, 1993). Even the very large altitudinal variation in sites was not evident in the ordination.

Table 1 List of taxa found in 6 habitat categories with number of records per habitat

Habitats	streams general	stream pools	stream riffles	residual pools	'canals'	wet walls	ponds	reservoirs
Sampling occasions	25	23	14	21	20	15	7	6
<i>Ablabesmyia longistyla</i> Ficc.	4	12	7	1	2	-	-	-
<i>Ablabesmyia</i> sp.	-	1	-	2	2	-	-	-
<i>Bryophaenocladus</i> sp.	-	-	-	-	1	-	-	-
♂ <i>Cardiocladius capucinus?</i> (Zett.)	-	-	-	-	1	-	-	-
♂ <i>Cardiocladius</i> sp.	-	-	-	-	1	-	-	-
♂ <i>Chaetocladius</i> sp.	-	-	-	1	-	-	-	-
♂ <i>Chironomus</i> sp.	2	9	2	10	4	-	4	5
♂ <i>Cladotanytarsus</i> sp.	-	2	1	-	-	1	-	-
♂ <i>Corynoneura</i> sp. A.	-	-	4	-	-	1	-	-
♂ <i>Cricotopus</i> (C.) <i>vierriensis</i> Goetgh.	2	-	-	-	6	1	-	-
♂ <i>Cricotopus</i> (L.) <i>ornatus</i> Mg.	-	-	-	-	-	-	-	1
♂ <i>Cricotopus</i> (L.) <i>sylvestris</i> Fabr.	-	1	-	-	-	-	1	-
♂ <i>Cricotopus</i> sp.	1	7	5	-	3	-	-	-
♂ <i>Dicrotendipes</i> sp.	-	2	-	1	-	-	2	-
♂ <i>Eukiefferiella ? ilikleyensis</i> (Edw.)	1	1	4	-	1	-	-	-
♂ <i>Eukiefferiella ? minor</i> (Edw.)	5	2	3	-	-	1	-	-
♂ <i>Halocladius</i> sp.	-	1	-	-	-	-	-	-
♂ <i>Limnophyes ? minimus</i> (Mg.)	1	-	-	-	-	-	-	-
♂ <i>Limnophyes</i> sp.	1	2	1	-	-	1	-	-
♂ <i>Macropelopia nebulosa</i> (Mg.)	1	5	1	-	-	-	-	-
♂ <i>Macropelopia</i> sp.	3	-	-	-	-	-	-	-
♂ <i>Metriocnemus ? fuscipes</i> (Mg.)	-	1	-	-	-	-	-	-
♂ <i>Metriocnemus obscuripes</i> (Holmgr.)	-	1	-	-	-	-	-	-
♂ <i>Metriocnemus</i> sp.	1	-	-	-	-	-	-	-
♂ <i>Micropsectra ? notescens</i> (Walk.)	1	3	1	-	-	-	-	-
♂ <i>Micropsectra</i> sp.	1	2	1	-	-	1	-	-
♂ <i>Orthocladius ? (Eudactylocladius) sp.</i>	2	-	-	-	1	2	-	-
♂ <i>Orthocladius (Euorthocladius) rivicola</i> Kieff	2	4	5	-	-	-	-	-
♂ <i>Orthocladius (Orthocladius) sp.</i>	-	4	5	1	1	-	-	-
♂ <i>Orthocladius</i> sp.	1	-	-	-	1	-	-	-
♂ <i>Orthocladius/Cricotopus</i> sp.	5	-	-	2	2	-	-	-
♂ <i>Parakiefferiella</i> sp.	1	-	-	-	-	-	-	-
♂ <i>Paramerina vaillanti</i> (Fitt.)	-	1	1	-	-	-	-	-
♂ <i>Paramerina mauritanica</i> (Firt.)	1	-	-	-	-	-	-	-
♂ <i>Paramerina</i> sp. Pel (sensu Langton 1991)	-	5	3	-	-	-	-	-
♂ <i>Paramerina</i> sp. 1	-	-	1	-	-	-	-	-
♂ <i>Parametriocnemus stylatus</i> (K.)	4	5	7	1	1	1	-	-
♂ <i>Paratrichocladius rufiventris</i> (Mg.)	3	4	7	1	6	2	-	-
♂ <i>Paratrichocladius</i> sp.	7	1	-	-	12	-	-	-
♂ <i>Phaenopsectra</i> sp.	3	6	1	3	2	-	-	-
♂ <i>Polypedilum</i> sp.	5	6	1	3	2	-	-	4
♂ <i>Procladius (Holotanypus) choreus</i> (Mg.)	1	4	-	1	-	-	1	-
♂ <i>Procladius (Psilotanypus) sp.</i>	-	-	-	-	1	-	-	-
♂ <i>Procladius</i> sp.	-	-	-	1	-	-	-	1
♂ <i>Psectrocladius limbatellus</i> Holmgr.	1	2	2	2	1	-	1	-
♂ <i>Psectrocladius ? octomaculatus</i> Wülk.	-	-	-	-	-	-	-	1
♂ <i>Psectrocladius</i> sp.	-	-	-	-	-	-	2	2
♂ <i>Rheocricotopus atripes</i> K.	4	4	4	1	-	-	-	-
♂ <i>Rheocricotopus</i> sp.	2	3	4	2	1	3	-	-

Table 1. cont.

Habitats	streams general	stream pools	stream riffles	residual pools	'canals'	wet walls	ponds	reservoirs
Sampling occasions	25	23	14	21	20	15	7	8
— <i>Rheotanytarsus</i> sp.	3	4	11	1	10			
— <i>Stictochironomus</i> sp.	1	1		3		-1		
— <i>Tanytarsus</i> sp.		7	2	1				
— <i>Thienemanniella</i> ? <i>clavicornis</i> K.		5	8					
— <i>Thienemanniella</i> sp.	2			1	2	1		
— <i>Thienemannimyia</i> sp.	1							
— <i>Trissopelopia</i> ? <i>flavida</i> (K.)	1	1	1					
● <i>Virgatanytarsus albisutus</i> (Santos Abreu)	1	4	6					
— <i>Zavelimyia</i> ? <i>nubila</i> (Mg.)	4	11	7	5	1			
— <i>Zavelimyia</i> sp.	4	1		5	1			

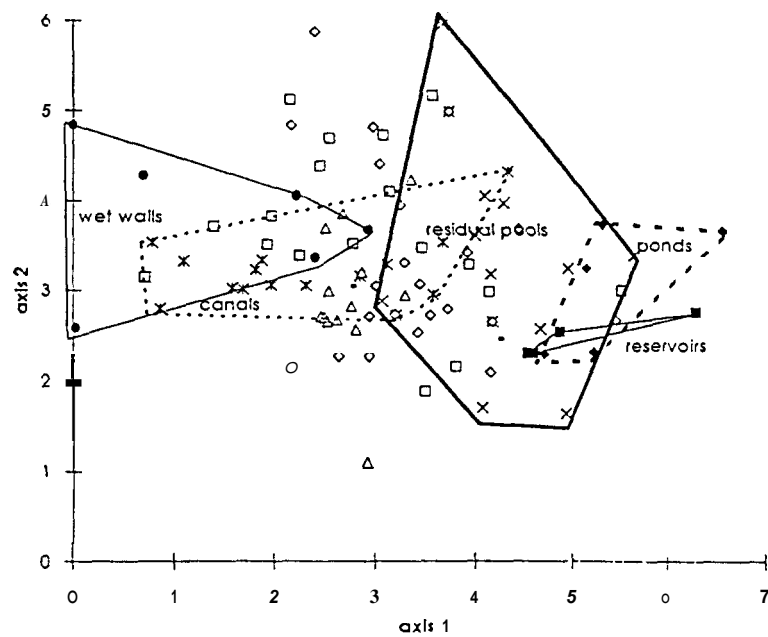


Figure 3n. Plots of ordination sample scores derived from DECORANA analysis of the entire sample site matrix. The polygons surround the data points in particular habitat types; (open square=streams general, open diamond=stream pools, open triangle=stream riffles, X=residual pools, closed circle=wet walls and seepages, closed square=reservoirs, closed diamond=ponds, = 'canals' (artificial channels), =light trap)

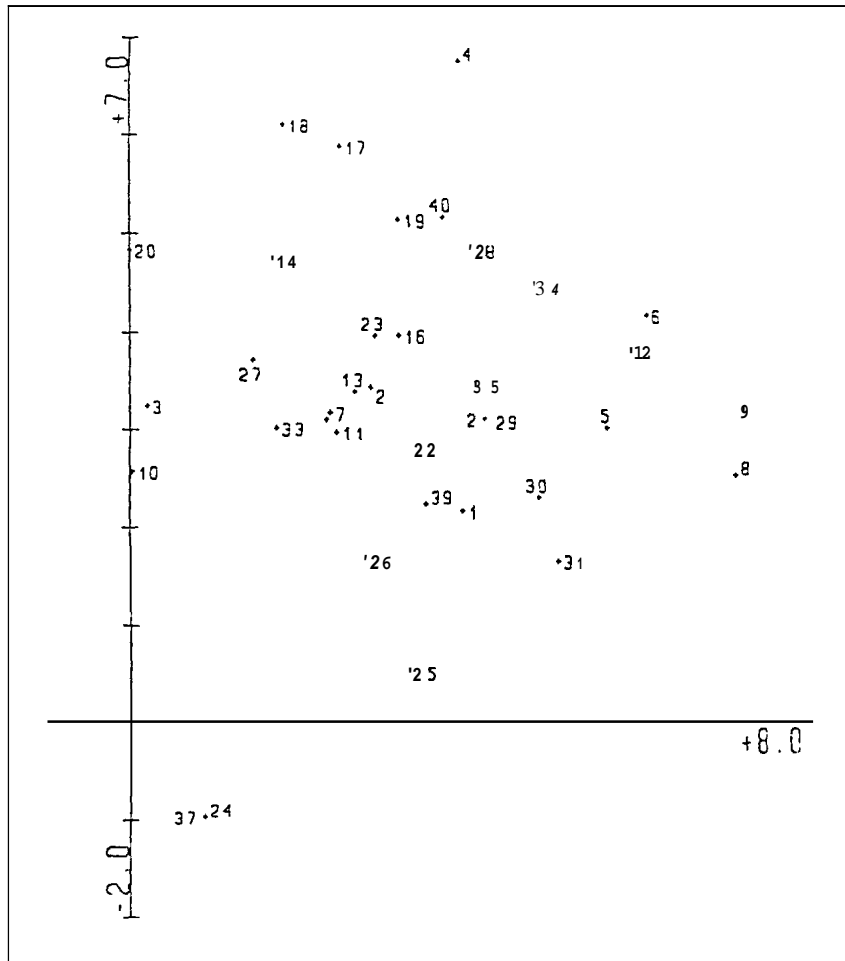


Figure 3b. Plots of ordination species scores. (axis1=x axis, axis2=y axis). 1 *Ablabesmyia* sp., 2 *Bryophaenocladus* sp., 3 *Cardiocladius* sp., 4 *Chaetocladus* sp., 5 *Chironomiu* spp., 6 *Cladotanytarsus* sp., 7 *Corynonettra* sp., 8 *Cricotopus ornatus*, 9 *C. sylvestris*, 10 *C. vierriensis*, 11 *Cricotopus* sp., 12 *Dicotendipes* sp., 13 *Eukiefferiella ?ilkleyensis*, 14 *E. ?minor*, 16 *Limnophyes* sp., 17 *Macropelopia* sp., 18 *Metriocnemus* spp., 19 *Micropsectra* sp., 20 *Orthocladus (Eudactylocladius)* sp., 21 *O.(Euorthocladus)* sp., 22 *O.(Orthocladus)* sp., 23 *Orthocladus/Cricotopus* spp, 24 *Parakiefferiella* sp., 25 *Paramerina* spp., 26 *Parametriocnemus stylatus*, 27 *Paratrithocladus rufiventris?*, 28 *Phaenopsectra* sp. 29 *Polypedilum* sp., 30 *Procladius* spp. 31 *Psectrocladius* spp., 32 *Rheocricotopus* sp., 33 *Rheotanytarsus* spp., 34 *Stictochironomus* sp., 35 *Tanytarsus* sp., 36 *Thienemanniella* sp., 37 *Thienemannimyia* group, 38 *Trissopelopia* sp. 39 *Virgatanytarsus* sp. 40 *Zavrelimyia* sp.

The relative richness of the habitat types is illustrated in Fig. 4. In the upper part of the figure it is clear that there is a positive relationship between total number of taxa and the number of sampling occasions, with the

exception that stream riffles have slightly higher number of taxa than expected and wet walls a lower number than expected. This point is borne out when the mean number of taxa per sample is considered (lower

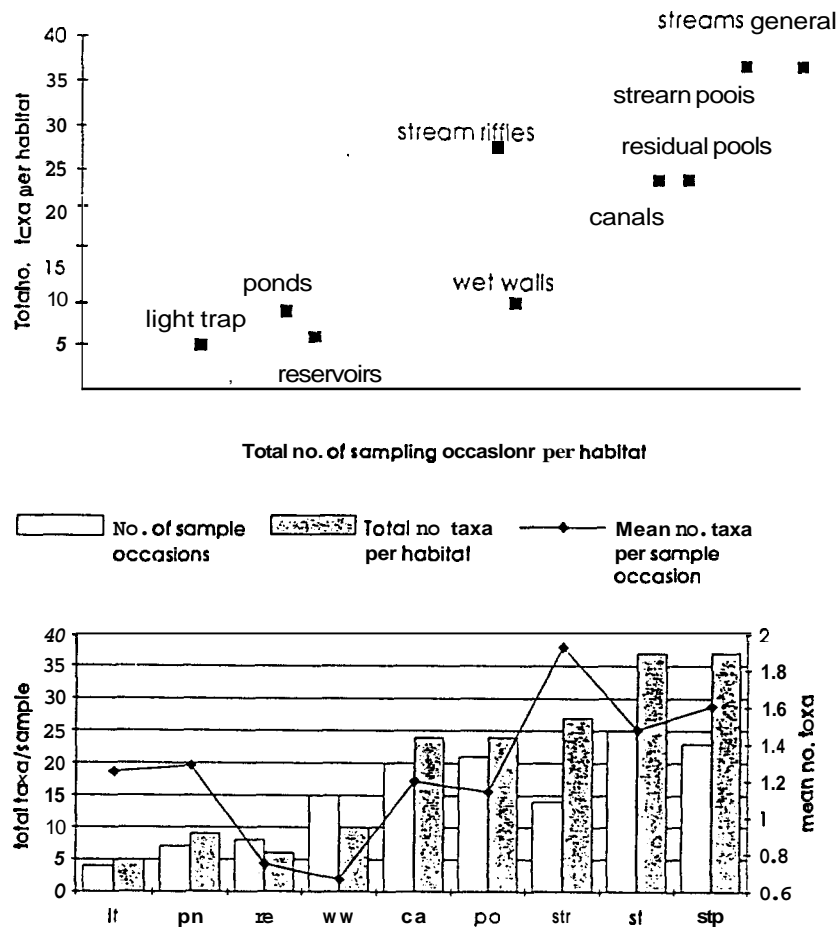


Figure 4. Taxon richness per sample effort (upper) and per habitat type (lower). (lt - light trap, pn - ponds, re - reservoirs, ww - wet walls, ca - canals, po - residual pools, and streams - general, st, riffles str and pools stp).

section of Fig. 4). The stream habitats contain more taxa than standing waters and within the latter ponds are richer than reservoirs.

DISCUSSION

The study of Chironomidae of Tenerife is still at an early stage. Although the work of Malmqvist *et al.* (1993) was systematic and detailed, collections of larval material do not usually provide the information

necessary for species identifications. Adult material in good condition is required.

Some taxa that were represented by adult, pupal and larval material were still difficult to identify to species. The genus *Paratrichocladius* definitely contained *P. rufiventris* but also included some forms which did not readily fit that species, with pupal euvia near *rufiventris* but with a point patch on tergite VII and adult features (gonocostic lobe) which did not fit *P. rufiventris*. Within the *Rheotanytarsus* complex there are probably three species.

One of these is new (*Rheotanytarsus* Pe 2, sensu Langton (1991) and is currently being described from all life-history stages (collected by PDA in 1983 and 1985). No pupal or adult material is available for the other *Rheotanytarsus* species. All queried species require more adult material to be certain of their identity.

The paucity of taxa found in this present survey may reflect not only the isolated and arid nature of much of the island but also the absence of suitable material which would have permitted further identification of the genera recorded. In neighbouring Morocco, Azzouzi and Laville (1987) recorded 65 species based on data from the literature between 1955 and 1986; with more collections this total rose to 134 species. Subsequent studies in lotic habitats (Azzouzi et al., 1992) have now raised this number to 223 species. In Tenerife a large increase in the recorded species can also be expected with more systematic study, despite the limited avail-

ability of freshwater habitat. Standing water habitats in the form of small reservoirs are increasing and it is likely that taxa lists from these habitats will increase over the next few years. Omitted from this study are terrestrial and marine habitats which can be expected to contribute a number of species (Arricape, 1987; Armitage and Tuiskunen, 1988).

It is too early for biogeographic speculation and this should await the availability of comprehensive species lists based largely on adult material.

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